Reference manual
Guidelines for otter spraint based diet analysis: searching, preservation, preparation and determination

Leeuwarden
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This manual is commissioned by:

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Abstract

This reference manual contains a simple standard method for finding and handling of otter (*Lutra lutra*) spraints for diet related research. The method describes, how to do spraint collection, how to preserve spraints, how to prepare spraints and how to determine to which species the prey remains belong. Also a description of the necessary materials is given. The reference manual also includes photographs of prey remains of several species that can be found in the otter spraints. The method is based on the analysis of 78 spraints from river and marsh habitats in the Netherlands. We anticipate that the manual will support and promote research on otter diet by amateurs interested in public science.
Preface

This reference manual is written as a thesis project of Hermsen, J. and Maarseveen, A., van for the BSc Animal Management, major Wildlife Management at the Van Hall Larenstein University of Applied Sciences in Leeuwarden. The project was supervised by Dr Strijkstra, A., and Mrs Heijer, M., den The research was commissioned by Drs. Niewold, F. J. J. of the Niewold Wildlife Infocentre and aimed for production of a realistic reference manual for prey remains in otter spraints, including a simple but effective standard method for the research on otter diet for amateur research in the Netherlands.

Leeuwarden, January 2012

Jessica Hermsen and Aloïse van Maarseveen
Contents
1. Introduction ................................................................. 5
2. Terminology ............................................................... 6
3. Standard method .......................................................... 7
   3.1 Material .................................................................. 7
   3.2 Flow chart; Field work; ............................................. 8
   3.3 Spraint preservation .................................................. 17
   3.4 Flow chart; spraint preparation; ................................... 18
   3.5 Flow chart; determination; ........................................... 19
4. Analysis ........................................................................ 20
   4.1 European perch ....................................................... 20
   4.2 Bream ................................................................... 21
   4.3 Silver bream ............................................................ 22
   4.4 Pike .................................................................... 23
   4.5 Tench .................................................................. 24
   4.6 Rudd ................................................................... 25
   4.7 Common roach ........................................................ 26
   4.8 Common carp .......................................................... 27
   4.9 Gudgeon ................................................................ 28
   4.10 Round goby ............................................................ 29
   4.11 Kessler’s goby ........................................................ 30
5. Reference cards ............................................................ 31
   5.1 Reference card; scale ................................................ 32
   5.2 Reference card; otolith .............................................. 34
Acknowledgements .................................................................. 36
References ........................................................................... 37
Articles .............................................................................. 37
CD-ROM; ........................................................................ 37
Internet ............................................................................. 37
Specialists ......................................................................... 37
1. Introduction

Not many diet studies on the Dutch otter (*Lutra lutra*) population have been done. Here for there is no standard method on how to do this research. The benefit of an easy standard method and reference manual may be that it promotes more research on the diet of this species by amateurs. This is the target group of this reference manual.

The method is based on information from literature study on otter diet studies, done in other countries, which was combined with expert opinions and our own practical experience on spraint analysis.

The manual explains the necessary steps for easy and efficient research on the otter’s diet based on spraints.

It starts off with necessary preparation for fieldwork and the materials needed. Furthermore a guide on how to do the field work is given, followed by a full explanation of how to gather, store, prepare and analyse the spraints for prey remains, and how to examine prey remains to determine the species.

The manual includes reference photographs, which can be used to compare prey remains too. The photographs are all made on prey remains found in otter spraints from otter of river and marsh habitats on several Dutch locations, and from boiled fish.
2. Terminology

Some terms are used which could be interpreted in different ways. To prevent confusion, some of these terms are defined below.

Table 1; Terminology, the right way of interpretation of several terms used in this reference manual.

<table>
<thead>
<tr>
<th>Term</th>
<th>Correct interpretation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Reference manual</td>
<td>A manual that contains pictures, photographs and drawing of the identifiable parts in the spraints. Prey remain in following research can be compared with these images in the manual.</td>
</tr>
<tr>
<td>Spraint</td>
<td>A small faecal secretion from an otter which contains prey remains, to mark their territory.</td>
</tr>
<tr>
<td>Prey remains</td>
<td>The (mostly hard) indigestible parts in a spraint, such as bones, fish scales, jawbones, vertebrae, hair, feathers.</td>
</tr>
<tr>
<td>Determination</td>
<td>The identification of a prey part, to its species.</td>
</tr>
<tr>
<td>Detergent</td>
<td>Washing fluid with in this case a solution of Sodium Carbonate, the product that is used for cleaning up spraints.</td>
</tr>
</tbody>
</table>
3. Standard method
This method is set up to make a study on the diet of the otter less time consuming.

3.1 Material
The following material is preferred in order to perform an optimal research (see table 2).

Table 2 Materials needed to perform this research about the diet of the Eurasian otter

<table>
<thead>
<tr>
<th>Specific for this research</th>
<th>General materials</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sieve 0.6mm</td>
<td>Measuring cup and stirrer</td>
</tr>
<tr>
<td>Sodium carbonate Na₂CO₃, 0.07%</td>
<td>Cutting board</td>
</tr>
<tr>
<td>Tissues</td>
<td>Waterproof marker and pen</td>
</tr>
<tr>
<td>Petri-dishes</td>
<td>Computer and printer</td>
</tr>
<tr>
<td>Microscope, (used for this reference manual; Olympus CHS)</td>
<td>Drawing material; pencil</td>
</tr>
<tr>
<td>Microscope slides, cover slips and demineralised water</td>
<td>Disposable gloves</td>
</tr>
<tr>
<td>Measuring scale (with accuracy of 0.1 gram)</td>
<td>Beaker</td>
</tr>
<tr>
<td>Cooking plate</td>
<td>Knife</td>
</tr>
<tr>
<td>Binocular-camera (used for this reference manual; Olympus SZ-CTV)</td>
<td>Stickers, that fit on the jars filled with ethanol</td>
</tr>
<tr>
<td>Tweezers</td>
<td>White paper sheets; to empty the sieve with the prey remains on</td>
</tr>
<tr>
<td>Spraints</td>
<td>Freezer</td>
</tr>
</tbody>
</table>

The determination books in table 3 were used during this research.

Table 3 Determination books needed to perform this research about the diet of the Eurasian otter in several study sites in the Netherlands; Wieden, Weerribben, Lindevallei, Rottige Meenthe, Doesburg, Giesbeek, and one in Germany; Peene.

<table>
<thead>
<tr>
<th>Determination book; title</th>
<th>Author</th>
<th>ISBN</th>
</tr>
</thead>
<tbody>
<tr>
<td>Atlas van schubben en andere beenachtige structuren van niet-zalmachtige zoetwatervissen, Organisatie ter verbetering van de Binnenvisserij;</td>
<td>Steinmetz, B. &amp; Muller, R. 1991</td>
<td>9789080012042</td>
</tr>
<tr>
<td>The Tailless Batrachians of Europe</td>
<td>Boulenger, G.A. 1897</td>
<td>9780548217207</td>
</tr>
<tr>
<td>Voedselenecologie van de ‘verwildere’ kat</td>
<td>Honstede, B. van &amp; Schut, G. 2010</td>
<td>594000</td>
</tr>
</tbody>
</table>
3.2 Flow chart; Field work; The steps to follow when you are going to search for spraints in the field

Legend

- Step 1: Preparing
- Step 2: Handling
- Step 3: Preservation

1. Literature study the general biology of the otter
2. Read the manual
3. Collection material needed in the field
4. Going to the study area and look for possible spraint locations
5. Prepare the material; print the field forms, fill the jars with ethanol
6. Look for spraints and determine the type of the spraint
7. Put the spraint in a plastic bag or a jar filled with ethanol and number them
8. Write down the information needed on the form provided
9. Put the spraints in the freezer until further examination
Field manual
This field manual contains guidance about how to collect spraints.

Preparation for a field trip
Collect all the materials mentioned in table 4. Make sure the stickers will fit on the jars and the jars are pre-filled with ethanol.

Print and cut out the labels designed in appendix 1 according to the amount of spraints that you anticipate to collect. These labels contain all important information that needs to be written down during field work.

Print some extra forms, don’t cut them; these will be used for the fresh spraints that are stored in jars with ethanol. You can number the jars with a pencil; these numbers refer to the ones you write down on your form.

Collect information on the research area and get a detailed map Google Earth, or 1:25000 map. Make an appointment with a specialist in this certain area to get information about the area and the otters that may live here.

Before starting check the weather conditions, this will influence the quality of the spraints. Rain: the spraints might get soak or flushed away. Cold weather close to freezing: DNA may be preserved for a longer time. Hot weather (20C+): the spraint will dry faster.

Table 4 Material list with the materials needed to perform this research about the diet of the Eurasian otter

<table>
<thead>
<tr>
<th>Kind of material</th>
<th>do of</th>
</tr>
</thead>
<tbody>
<tr>
<td>Clipboard</td>
<td></td>
</tr>
<tr>
<td>Eraser</td>
<td></td>
</tr>
<tr>
<td>GPS</td>
<td></td>
</tr>
<tr>
<td>Pencil</td>
<td></td>
</tr>
<tr>
<td>Plastic bags to collect the spraints</td>
<td></td>
</tr>
<tr>
<td>Plastic jars (with screw caps) 150 ml</td>
<td></td>
</tr>
<tr>
<td>Print paper (to make labels)</td>
<td></td>
</tr>
<tr>
<td>Stickers to put on the jars</td>
<td></td>
</tr>
<tr>
<td>Ethanol 100%, 70%</td>
<td></td>
</tr>
</tbody>
</table>

Fieldwork
Otters have typical places to spraint, where the water is entered or exited and on conspicuous spots around passages where the otter often walks through. Places to look for spraints are indicated in table 5 and illustrated with photographs. You might also invite sprainting by creating a spot.

Table 5 Places to look for otter spraints in the field, with their explanation.

<table>
<thead>
<tr>
<th>Place</th>
<th>Explanation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Otter passages (figure 1 and 2, appendix 2)</td>
<td>A small hole in the scrubs with a section of about 30 cm</td>
</tr>
<tr>
<td>Otter path (figure 1, appendix 2)</td>
<td>A clear path, where the vegetation has often been walked on, comes out in these holes and goes to another place. Often from one water place to the other</td>
</tr>
<tr>
<td>Water edge</td>
<td>Look for spots at the water edge where the vegetation is flat (walked on) or brown/ yellow (urinated) or where there is an object available to spraint on, like a stone. Drove outs for game are a good spot to look.</td>
</tr>
</tbody>
</table>
Create a spot where otters often come, by offering them a tree trunk or another object.

Look for traces of the otter at the place where the spraint was found (see figure 3). This will increase the determination reliability that the spraint originates from an otter. Write down where the spraint was found (shade, sun, under a bridge, etc.) as this might have influenced the quality of the spraint and must be taken into account for further research.

There are different types of spraints, (table 6). The type needs to be determined during collection to know if the spraint contains DNA. Jellies contain most DNA, as a jelly is epithelia mucus from the intestine. Fresh spraints contain usable DNA as well.

**Table 6 Spraint types with figure numbers for examples, and their explanation.**

<table>
<thead>
<tr>
<th>Spraint</th>
<th>Figure</th>
<th>Explanation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Jelly</td>
<td>Figure 7 and 8, appendix 2</td>
<td>Mucus, yellow, white or green</td>
</tr>
<tr>
<td>Fresh spraint</td>
<td>Figure 6, appendix 2</td>
<td>Wet, strong smell, soft when you poke in it</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(with a stick)</td>
</tr>
<tr>
<td>Old spraint</td>
<td>Figure 5, appendix 2</td>
<td>Dry, smell is not so strong, hard</td>
</tr>
</tbody>
</table>

When the origin of the spraint is unclear the smell may be helpful. Otter spraints have a strong fish smell. This combined with knowledge about the area, the otter and traces found at the location will determine if it safe to assume that it is an otter spraint.

Old spraints are collected in a plastic bag. The information on the label should be filled out as complete as possible and is put in second plastic bag together with plastic bag with the spraint. This prevents the label from getting wet and unreadable. Make sure to use capable letters, this makes sure that everyone can read it. Process the data on a computer as soon as possible after the fieldtrip. Fresh spraints can be used for DNA research and when used for that potential purpose, need to be stored in 70% ethanol. Number the jars with the fresh spraints and link these to a paper or digital archive, i.e. a form or a date file.

**Finishing spraint collection**
The collected spraints need to be stored frozen at -20C until they are used for further examination.
Figure 1 Passage and path of the otter (photographed at Wieden/Weerribben on 22 November 2011)

Figure 2 Otters passage (photographed at Wieden/Weerribben on 22 November 2011)
Figure 3 Scratch traces of an otter (photographed at Wieden/Weerribben on 22 November 2011)

Figure 4 Otter spraint (photographed at Wieden/Weerribben on 22 November 2011)
Figure 5 Otter spraint (photographed at Wieden/Weerribben on 22 November 2011)

Figure 6 Otter spraint (photographed at Wieden/Weerribben on 22 November 2011)
Figure 7 Otter spraint, jelly (photographed at Wieden/Weerribben on 22 November 2011)

Figure 8 Otter spraint, jelly, photographed by Bosma, H. (2011)
3.3 Spraint preservation

3.4 Flow chart; spraint preparation; the steps to follow when you clean the spraints (remove waste and mucus from the prey remains)

Legend

- **Step 1**: Prepare the spraints
- **Step 2**: Rinse the prey remains
- **Step 3**: Dry the prey remains

**Make a 0.07% solution of Na₂CO₃ and water**

**Leave the spraint soaking in the solution for a week.** Put each spraint individually in a beaker.

**Sieve the solution. Use a 0.6mm mesh.**

**Rinse the prey remains with tap water, in the sieve.**

**Pad dry the sieve at the bottom with tissues.**

**Empty the sieve on a clean paper sheet. (Knocking the sieve on the sheet to get the remains out)**

**Number the petri-dish and put the remains in it, leave the petri-dish open.**

**Air dry the remains for a week, then close the petri-dish till further examination.**

Air dry the remains for a week, then close the petri-dish till further examination.
3.5 Flow chart; determination; the steps to follow when you want to determine the species of the prey remains

Legend
- **Blue**: Step 1: Preparing
- **Green**: Step 2: Handling
- **Red**: Step 3: Data

**Step 1: Preparing**
- Take all the otoliths, bones, hair/feather s and other large prey remains out

**Step 2: Handling**
- Take a scale at random (in the middle of the petri-dish)
- Put the scales on a slide for under the microscope
- Examine them under the microscope
- Determine the prey species with the help of the determination books
- Measure the size of the otoliths and calculate the fish size and mass (with the use of the CD-ROM otoliths of the North Sea fish, fish identification key by means of otoliths and other hard parts by Leopold, M., et al. 2001).
- Put them under the binocular camera and make pictures
- Make a database from the photo’s
4. Analysis

Determination material for the European perch, Bream, Silver bream, Pike, Tench, Rudd, Common roach, Common carp, Gudgeon, Round goby and the Kessler’s goby is given. These species were encouraged after the analysis of 78 spraints of otters in Dutch waters.

4.1 European perch

Table 7 Determination material for the European perch (*Perca fluviatilis*)

<table>
<thead>
<tr>
<th>Scientific name</th>
<th><em>Perca fluviatilis</em></th>
</tr>
</thead>
<tbody>
<tr>
<td>Exterior</td>
<td>Figure 9 (Sportvisserij Nederland, 2012)</td>
</tr>
<tr>
<td>Scale</td>
<td>Figure 10</td>
</tr>
<tr>
<td>Otolith</td>
<td>Figure 11</td>
</tr>
<tr>
<td>Spine</td>
<td>Figure 12</td>
</tr>
</tbody>
</table>

Figure 9 *Perca fluviatilis* (Sportvisserij, 2012); 1; The Perch has two separated dorsal fins of which only the first one has hard spines. 2; The Perch has a back spot on the back-side of the first dorsal fin. 3; The Perch has vertical dark ties across the body.

Figure 10 *Perca fluviatilis* scale, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4

Figure 11 *Perca fluviatilis* otolith, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4

Figure 12 *Perca fluviatilis* spine, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 0.67
4.2 Bream

Table 8 Determination material for the Bream (Abramis brama)

<table>
<thead>
<tr>
<th>Scientific name</th>
<th>Abramis brama</th>
</tr>
</thead>
<tbody>
<tr>
<td>Exterior</td>
<td>Figure 13 (Sportvisserij Nederland, 2012)</td>
</tr>
<tr>
<td>Scale</td>
<td>Figure 14</td>
</tr>
<tr>
<td>Otolith</td>
<td>Figure 15</td>
</tr>
<tr>
<td>Jaw</td>
<td>Figure 16</td>
</tr>
<tr>
<td>Spine</td>
<td>Figure 17</td>
</tr>
</tbody>
</table>

Figure 13 Abramis brama (Sportvisserij, 2012); 1; Number of rows of scales above the side-line, counted from diagonal pointing arrow to the dorsal fin, contains 12-14 scales (excluded the one from the side-line itself). 2; The eye-diameter is smaller than the distance from the eye till the point of its mouth. 3; The mouth is pointing downwards and is far bulging.

Figure 14 Abramis brama scale, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.

Figure 15 Abramis brama otolith, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.

Figure 16 Abramis brama, jaw, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 0.67.

Figure 17 Abramis brama spine, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 0.67. Upper photo; side view, bottom photo; front view.
4.3 Silver bream

Table 9 Determination material for the Silver bream (*Blicca bjoerkna*)

<table>
<thead>
<tr>
<th>Scientific name</th>
</tr>
</thead>
<tbody>
<tr>
<td>Exterior</td>
</tr>
<tr>
<td>Scale</td>
</tr>
<tr>
<td>Otolith</td>
</tr>
<tr>
<td>Spine</td>
</tr>
<tr>
<td>Jaw</td>
</tr>
</tbody>
</table>

Figure 18 *Blicca bjoerkna* (Sportvisserij Nederland, 2012); 1; Number of rows of scales above the side-line, counted from the oblique pointed arrow to the dorsal fin, counts 8-10 (excluded is the scale on the side-line). 2; The eye-diameter is larger than the distance from the eye to the point of the mouth.

Figure 19 *Blicca bjoerkna* scale, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.

Figure 20 *Blicca bjoerkna* otolith, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.

Figure 21 *Blicca bjoerkna* spine, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 0.67.

Figure 22 *Blicca bjoerkna* jaw, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 0.67.
### 4.4 Pike

**Table 10 Determination material for the Pike (Esox lucius)**

<table>
<thead>
<tr>
<th>Scientific name</th>
<th>Esox lucius</th>
</tr>
</thead>
<tbody>
<tr>
<td>Exterior</td>
<td>Figure 23 (Sportvisserij Nederland, 2012)</td>
</tr>
<tr>
<td>Scale</td>
<td>Figure 24</td>
</tr>
<tr>
<td>Otolith</td>
<td>Figure 25</td>
</tr>
<tr>
<td>Spine</td>
<td>Figure 26</td>
</tr>
<tr>
<td>Jaw</td>
<td>Figure 27</td>
</tr>
</tbody>
</table>

![Figure 23](Esox lucius (Sportvisserij Nederland, 2012); 1: Anal fin and dorsal fin occurs far backwards on the body. 2: The head runs out into a flat, wide mouth. 3: All over the body there are gold-colored dots and stripes.)

![Figure 24](Esox lucius scale, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.)

![Figure 25](Esox lucius otolith, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.)

![Figure 26](Esox lucius spine, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 0.67.)

![Figure 27](Esox lucius jaw, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 0.67.)
4.5 Tench

Table 11 Determination material for the Tench (*Tinca tinca*)

<table>
<thead>
<tr>
<th>Scientific name</th>
<th>Tinca tinca</th>
</tr>
</thead>
<tbody>
<tr>
<td>Exterior</td>
<td>Figure 28 (Sportvisserij Nederland, 2012)</td>
</tr>
<tr>
<td>Scale</td>
<td>Figure 29</td>
</tr>
<tr>
<td>Otolith</td>
<td>Figure 30</td>
</tr>
<tr>
<td>Spine</td>
<td>Figure 31</td>
</tr>
</tbody>
</table>

Figure 28 *Tinca tinca* (Sportvisserij Nederland, 2012); 1: The iris is orange-colored. 2: The fins (point out is the dorsal fin) have a convex-shape. 3: There are two little mouth-wires present.

Figure 29 *Tinca tinca* scale, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.

Figure 30 *Tinca tinca* otolith, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.

Figure 31 *Tinca tinca* spine, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 0.67.
4.6 Rudd

Table 12 Determination material for the Rudd (*Scardinius erythrophthalmus*)

<table>
<thead>
<tr>
<th>Scientific name</th>
<th>Scardinius erythrophthalmus</th>
</tr>
</thead>
<tbody>
<tr>
<td>Exterior</td>
<td>Figure 32 (Sportvisserij Nederland, 2012)</td>
</tr>
<tr>
<td>Scale</td>
<td>Figure 33</td>
</tr>
<tr>
<td>Otolith</td>
<td>Figure 34</td>
</tr>
<tr>
<td>Spine</td>
<td>Figure 35</td>
</tr>
<tr>
<td>Jaw</td>
<td>Figure 36</td>
</tr>
</tbody>
</table>

Figure 32 *Scardinius erythrophthalmus* (Sportvisserij Nederland, 2012); 1; The mouth is pointing upwards. 2; The front-side of the dorsal fin is clearly further to the tail of the fish than the pelvic fins.

Figure 33 *Scardinius erythrophthalmus* scale, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.

Figure 34 *Scardinius erythrophthalmus* otolith, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.

Figure 35 *Scardinius erythrophthalmus* spine, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 0.67.

Figure 36 *Scardinius erythrophthalmus* photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 0.67.
### 4.7 Common roach

Table 13 Determination material for the Common roach (*Rutilus rutilus*)

<table>
<thead>
<tr>
<th>Scientific name</th>
<th><em>Rutilus rutilus</em></th>
</tr>
</thead>
<tbody>
<tr>
<td>Exterior</td>
<td>Figure 37 (Sportvisserij Nederland, 2012)</td>
</tr>
<tr>
<td>Scale</td>
<td>Figure 38</td>
</tr>
<tr>
<td>Otolith</td>
<td>Figure 39</td>
</tr>
</tbody>
</table>

Figure 37 *Rutilus rutilus* (Sportvisserij Nederland, 2012); 1; The mouth is pointing forwards. 2; In the top of the eye, a red spot is shown. 3; The front-side of the dorsal fin is just as far from the tail as the pelvic fins.

Figure 38 *Rutilus rutilus* scale, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.

Figure 39 *Rutilus rutilus* otolith, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.
4.8 Common carp

Table 14 Determination material for the Common carp (Cyprinus carpio)

<table>
<thead>
<tr>
<th>Scientific name</th>
<th>Cyprinus carpio</th>
</tr>
</thead>
<tbody>
<tr>
<td>Exterior</td>
<td>Figure 40 (Sportvisserij Nederland, 2012)</td>
</tr>
<tr>
<td>Scale</td>
<td>Figure 41</td>
</tr>
<tr>
<td>Otolith</td>
<td>Figure 42</td>
</tr>
</tbody>
</table>

Figure 40 Cyprinus carpio (Sportvisserij Nederland, 2012); 1; There are four mouth-wires present, from which two in de corners of the mouth and two shorter ones on top of the upper lip. 2; The edge of the large dorsal fin is hollow incised. 3; The first fin rays of the dorsal fin is firm serrated.

Figure 41 Cyprinus carpio scale, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.

Figure 42 Cyprinus carpio otolith, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.
4.9 Gudgeon

Table 15 Determination material for the Gudgeon (*Gobio gobio gobio*)

<table>
<thead>
<tr>
<th>Scientific name</th>
<th><em>Gobio gobio gobio</em></th>
</tr>
</thead>
<tbody>
<tr>
<td>Exterior</td>
<td>Figure 43 (Sportvisserij Nederland, 2012)</td>
</tr>
<tr>
<td>Scale</td>
<td>Figure 44</td>
</tr>
<tr>
<td>Otolith</td>
<td>Figure 45</td>
</tr>
</tbody>
</table>

Figure 43 *Gobio gobio gobio* (Sportvisserij Nederland, 2012); 1; The mouth is pointing downwards. 2; There are two mouth-wires present, one in each corner of the mouth.

Figure 44 *Gobio gobio gobio* scale, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.

Figure 45 *Gobio gobio gobio* otolith, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.
4.10 Round goby

Table 16 Determination material for the Round goby (*Neogobius melanostomus*)

<table>
<thead>
<tr>
<th>Scientific name</th>
<th><em>Neogobius melanostomus</em></th>
</tr>
</thead>
<tbody>
<tr>
<td>Exterior</td>
<td>Figure 46 (Sportvisserij Nederland, 2012)</td>
</tr>
<tr>
<td>Scale</td>
<td>Figure 47</td>
</tr>
</tbody>
</table>

Figure 46 *Neogobius melanostomus* (Sportvisserij Nederland, 2012) 1; The eyes are highly placed into the head. 2; The pelvic fins are fused to a suction-plate, with which the fish can suck itself on a stone-based underground. 3; This species is yellow-grey-colored and has a checker-pattern. 4; In the back of the first dorsal fin, a clear black spot is shown. Males are black of colour during mating season.

Figure 47 *Neogobius melanostomus* scale, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.
4.11 Kessler’s goby

Table 17 Determination material for the Kessler’s goby (*Neogobius kesslerii*)

<table>
<thead>
<tr>
<th>Scientific name</th>
<th><em>Neogobius kesslerii</em></th>
</tr>
</thead>
<tbody>
<tr>
<td>Exterior</td>
<td>Figure 48 (Sportvisserij Nederland, 2012)</td>
</tr>
<tr>
<td>Scale</td>
<td>Figure 49</td>
</tr>
</tbody>
</table>

Figure 48 *Neogobius kesslerii* (Sportvisserij Nederland, 2012); 1; The eyes are close together and highly placed into the head. A big, wide head with swollen cheeks and lips, and a mouth pointing upwards. A neck with scales. 2; Pelvic fins are fused with which the fish can suck itself on a hard-based underground. 3; The body and the head are red-brown marbled. 4; The bases of both dorsal fins touch each other. Dorsal fins with horizontal red-brown ties on a lighter underground and without a black spot.

Figure 49 *Neogobius kesslerii* scale, photographed with a binocular-camera (Olympus SZ-CTV) with an enlargement of 1.4.
5. Reference cards

The following cards are developed to help determine the species from an otolith or scale found. By answering the questions you will find the species the scale or otolith belongs to.
5.1 Reference card; scale

- Dorsal edge
- Ventral edge
- Anterior margin
- Chromatophore in derms
- Ctenii
- Radii
- Circuli
- Focus
- Ventral edge
Is the scale oval or round?

**Oval:**
- Does the scale have Ctenii?
  - Yes:
    - Does the scale have Chromatophore in derms?
      - Yes: Pike (Esox lucius)
      - No: Tench (Tinca tinca)
  - No: Round goby (Neogobius melanostomus)

**Round:**
- Does the scale have Radii?
  - Yes:
    - Does the scale have Ctenii?
      - Yes:
        - Does the scale have round Anterior margins?
          - Yes: European perch (Perca fluviatilis)
          - No: Bream (Abramis brama)
      - No: Does the scale have a curvy edge?
        - Yes: Round goby (Neogobius melanostomus)
        - No: Rudd (Scardinius erythrophthalmus)
      - No: Round goby (Neogobius melanostomus)
  - No:
    - Does the scale has Radii at both edges?
      - Yes: Common roach (Rutilus rutilus)
      - No: Common carp (Cyprinus carpio)

Species:
- Tench (Tinca tinca)
- Pike (Esox lucius)
- Round goby (Neogobius melanostomus)
- Silver bream (Blicca bjoerkna)
- European perch (Perca fluviatilis)
- Kessler’s goby (Neogobius kessleri)
- Bream (Abramis brama)
- Rudd (Scardinius erythrophthalmus)
- Gudgeon (Gobio gobio)
- Common roach (Rutilus rutilus)
- Common carp (Cyprinus carpio)
5.2 Reference card; otolith

- Dorsal edge
- Antirostrum
- Excisura major
- Excisura minor
- Pararostrum
- Postrostrum
- Rostrum
- Ventral edge
Is the otolith round or oval?

Oval:
- Is the outline jagged?
  - Yes: Does the otolith have an Excisura minor
  - Yes: European perch (Perca fluviatilis)
  - No: Silver bream (Blicca bjoerkna)
- No: Does the otolith have an Excisura major
  - No: Silver bream (Blicca bjoerkna)
  - Yes: Tench (Tinca tinca)

Round:
- Is the outline curvy?
  - Yes: Tench (Tinca tinca)
  - No: Does the otolith have a weak inlet?
    - Yes: Rudd (Scardinius erythrophthalmus)
    - Yes: Common roach (Rutilus rutilus)
    - No: Bream (Abramis brama)
    - Yes: Common carp (Cyprinus carpio)
    - No: Bream (Abramis brama)
- Yes: Is the rostrum pointy or round
  - Yes: Gudgeon (Gobio gobio)
  - No: Bream (Abramis brama)
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CD-ROM;

Internet

Specialists
Bosma, H. (2011) Wetterskyp Fryslan